

# DISTRIBUTION OF CLOSTRIDIUM BOTULINUM IN CURED FISHERY PRODUCTS

by

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## ABSTRACT

A total of forty cured fish samples procured from local retail markets in and around Cochin were analysed for the moisture content, salt content, water activity ( $a_w$ ) and for the presence of *Clostridium botulinum*. Wide variations in the salt concentration and moisture level were observed in the cured fish samples examined. Fifty five percent (22/40) of the samples had water content in the range of 45-51%. The sodium chloride content of the cured fish samples varied between 16-25% in only 60 percent of the samples. The sodium chloride content of less than 7% was noticed in 8 prawn samples. The  $a_w$  of the samples varied greatly. *Clostridium botulinum* was detected in 13% of the cured fish samples and type D was predominant (4/5) followed by type C (1/5). *Clostridium botulinum* spores remain viable at  $a_w$  level 0.75. The incidence of *C. botulinum* in cured fish should emphasize the need for adequate brining and drying to protect these products until their final use.

## INTRODUCTION

Cured fish is the only traditional fish product of overall commercial importance in India accounting for utilization of about 30 percent of marine fish landings (Sripathy 1983). The process of curing fish imparts a degree of microbiological stability to the product but their stability is a function of several factors including the salt level reached after brining and the amount of heat applied. Since these factors vary from product to product, the microbiological quality and safety of cured fishery products vary.

*Clostridium botulinum* is one of the food safety hazards associated with fish. There are several botulism hazards associated with seafoods or fish (Sakaguchi 1979, Huss 1981, Dodds 1993). Even though the salt added during the production of cured fish is inhibitory to microbial growth, there have been outbreaks of botulism associated with salt dried fish (Sakaguchi 1979, Centre for Disease Control 1987, Hauchild 1989, Weber *et al.* 1993, McClure *et al.* 1994).

Incidence of *Clostridium botulinum* was reported in fish from India (Lalitha and Iyer 1990, Lalitha and Surendran 1992). The mere presence of *C. botulinum* on a food product will not cause illness, conditions much be such that viable *C. botulinum* spores are present and are given the opportunity to germinate and produce toxin. At present there is paucity of data on the occurrence of *C. botulinum* in cured fish. This paper describes a survey on the distribution of *C. botulinum* in cured fish on sale in markets in Cochin, India.

## MATERIALS AND METHODS

Cured fish/shellfish samples from the retail outlets, in and around Cochin, were aseptically collected in sterile polythene bags and transported to the laboratory for the estimation of chemical parameters such as salt content, moisture content and water activity and for the detection of *C. botulinum*.

Cured fish/shellfish samples procured from local retail outlets were analysed for sodium chloride and moisture contents. Moisture and sodium chloride contents were determined according to the methods

followed by AOAC (1990). The water activity of cured fish samples were estimated using a water activity meter (Lufft  $a_w$  - Wert-Messer, Germany).

Cured fishery products obtained from retail outlets in and around Cochin were examined for the presence of *C. botulinum*. A 3-5g sample of cured fish was inoculated into 25 ml Cooked Meat Medium (CMM) after removing dissolved oxygen by steaming 10-15 min and cooling quickly without agitation. Sterile paraffin oil was used to cover the surface. Inoculated tubes were incubated at 30°C for 6 days.

After incubation for 6 days at 30°C, cultures were centrifuged at 10000 x g and 4°C for 20 min (Remi, India) and each supernatant was adjusted to pH 6.2 with 1N HCl and frozen at -15°C overnight to eliminate non-specific mouse deaths (Baker *et al.* 1990). Supernatants were stored at -15°C until tested. Toxicity of the supernatant was tested in mice and the toxin type was identified using a procedure followed by the US Food and Drug Administration (Solomon *et al.* 1995).

## RESULTS AND DISCUSSION

A total of 40 cured fish/shellfish samples procured from local retail markets were examined for the presence of *C. botulinum* and the data are summarised in table 1 and 2. *Clostridium botulinum* was detected in 13% of the samples with type D predominating (4/5) followed by type C (1/5).

Table 1. Chemical characteristics of cured fish.

Common English name of sample	Scientific name	No. of samples examined	Moisture %		Salt %		$a_w$	
			Average	Range	Average	Range	Average	Range
White sardine	<i>Escualosa thoracata</i>	10	45.07	39-49	16.75	13-19	0.76	0.75-0.80
Malabar Tongue sole	<i>Cynoglossus macrostomus</i>	5	45.84	40-49	17.32	10-25	0.79	0.75-0.83
Ribbon fish	<i>Lepturacanthus savala</i>	7	46.13	37-51	17.10	15-21	0.76	0.75-0.80
Malabar anchovy	<i>Thryssa malabarica</i>	8	45.35	44-48	16.69	14-20	0.75	0.75-0.76
White bait	<i>Stolephorus bataviensis</i>	2	45.76	45-46	21.51	20-22	0.75	0.75
Prawn	<i>Metapenaeus dobsoni</i>	8	18.39	12-22	4.61	3-6	0.75	0.65-0.88

The water activity ( $a_w$ ) of samples varied greatly (Fig. 1). Eighty eight percent of the samples had  $a_w$  values in the range of 0.71-0.80 while 10% had  $a_w$  levels of 0.81 to 0.88. The analysis of the water content of the cured fishery products revealed that 55% (22/40) samples had water content in the range of 45-51% and in 25% of the samples, water content varied between 36-45%. In cured prawn samples, water content was still lower (<22%). Sodium chloride content of the cured fish samples varied between 16-25% in only 60 percent of the samples. 20% had sodium chloride content in the range of 11-15%. Sodium chloride content of less than 7% was noticed in 8 prawn samples in the present study. Wide variations in the salt concentration and moisture level were observed in the cured fish samples examined (Fig. 2 and 3).

Table 2. Occurrence of *Clostridium botulinum* in cured fish.

Sample	No. of samples examined	No. of samples positive for <i>C. botulinum</i>	<i>C. botulinum</i> Type identified
Escualosa thoracata	10	1	C
Cynoglossus macrostomus	5	1	D
Lepturacanthus savala	7	1	D
Thryssa malabarica	8	1	D
Stolephorus bataviensis	2	-	-
Metapenaeus dobsoni	8	1	D

Hauschild (1989) reviewed the prevalence of *C. botulinum* in fresh processed fish. Salted fish from the Caspian Sea were 29 percent positive while smoked fish carried spores in 1.1-20 percent of samples. Dodds (1993) reported incidence of *C. botulinum* in 63% of salted carp from the Caspian sea and type E spores were found at the level of <60/10g in salted fish from the Caspian sea and 490/10g in salted carp. A low incidence of *C. botulinum* (13%) was found in cured fish in the present study.

The viability of *C. botulinum* spores at water activity level 0.75 was shown in the present study by the toxic enrichment cultures of cured fish with water activity 0.75. The viability of *C. botulinum* spores for long periods in many foods of low water activity have been reported earlier (Troller and Christian 1978; Genigeorgis and Riemann 1979; ICMSF 1980). These spores are of great concern after rehydration of such foods (ICMSF 1980). There have been outbreaks of botulism associated with salt dried fish (Troller and Christian 1978; Sakaguchi 1979; Huss 1981; McClure *et al.*, 1994).

It is noted that 56% of the cured fish samples (18/32) had moisture content in the range of 46-51% and 44% (14/32) had moisture contents in the range of 37-45%, values higher than the Bureau of Indian Standards (BIS) (IS: 2882, 1964; IS: 5198, 1969 and IS: 2883, 1976) which prescribe a range of 10-35% moisture normally and 40-45% moisture in respect of certain big fish (Gopakumar and Devadasan 1983). Most of the samples do not conform to the BIS levels of salt (IS: 594, 1962). It is evident from the results of the present study that the majority of the cured fish at the retail level contained inadequate salt levels and high moisture levels as reported earlier (George Joseph *et al.*, 1986, 1988; Kalaimani *et al.*, 1988; Prasad *et al.*, 1994). Kalaimani *et al.*, (1988) reported  $a_w$  levels of less than 0.80 in 30 percent of the cured fish examined from Quilon, Vizhinjam and Tuticorin. In the present study nearly 90% of the samples had  $a_w$  values in the range of 0.71 to 0.80. Wide variations were also observed in the  $a_w$  levels of cured fish examined in the present study.

A survey of the cured fish sold at the retail markets in Cochin revealed wide variation in the salt concentration indicating inadequate control of the brining process in a number of products. Recently outbreaks of botulism from *Kapchunika* (salt-cured, air dried uneviscerated white fish) in USA and Israel (CDC 1987; Slater *et al.*, 1989; Telzak *et al.*, 1990) and from aseikh (uneviscerated fish) in Egypt (Weber *et al.*, 1993) have been reported and attributed to a poorly controlled salting process. *C. botulinum* types B and E were involved in the above outbreaks.

The predominance of *C. botulinum* types C and D in fresh fish have been reported from tropical areas (Tanasugarn 1979; Haq and Suhadi 1981; Lalitha and Surendran 1992). *C. botulinum* types C and D are the predominant types in sediment samples from Bangladesh, Indonesia, Thailand and India (Huss 1980; Haq and Suhadi 1981; Tanasugarn 1979; Lalitha and Surendran 1993) and soil samples from Indonesia and Thailand (Tanasugarn 1979; Hayashi *et al.*, 1981). The finding that types C and D are the predominant types in cured fish as shown by the present study is as expected.

*Clostridium botulinum* types C and D have been reported to cause intoxications only in animals (Smith 1990; Hauschild 1993). However, cases of human botulism due to types C and D have occurred earlier (Roberts and Gibson 1979; Hauschild 1993). Sonnabend and Sonnabend (1981) detected *C. botulinum* type D and toxin in patients who died unexpectedly with severe respiratory difficulty and dysphagia. Fastidiousness of type C in experimental and captive monkeys (Dolman *et al.*, 1961; Barnes and Mead 1986) and recent infant botulism by type C in Japan (Oguma *et al.*, 1990) indicate a potential risk. The possible hazard due to salted fish is highlighted by the incidents of botulism associated with it.

The present study indicates a low incidence of *C. botulinum* (13%) and predominance of type C and D in cured fishery products. *Clostridium botulinum* spores remain viable at water activity ( $a_w$ ) level 0.75 and are able to grow and produce toxin after rehydration of such foods. The detection of *C. botulinum* in cured fish indicate either post processing contamination or the ability of *C. botulinum* strains to survive the salting process during the manufacture of cured fish. Wide variations in the salt concentration of cured fishery products indicate inadequate control of the brining process. Therefore, the incidence of *C. botulinum* in 13 percent of the cured fishery products should emphasize the need for adequate brining and drying to protect these products until their final use. Proper hygienic conditions in the curing yards are to be encouraged for the production of good quality products.

## References

- AOAC. 1990. Official Methods of Analysis (Ed. Helrich, K.) 15th edition. Arlington, Virginia, U.S.A., AOAC Inc.
- Baker, D.A., Genigeorgis, C. and Garcia, G. 1990. Prevalence of *Clostridium botulinum* in seafood and significance of multiple incubation temperatures for determination of its presence and type in fresh retail fish. *J. Food Protect* 53(8), 668-73.
- Barnes, E.M. and Mead, G.C. 1986. *Anaerobic bacteria in habitats other than man*. Oxford, London, Blackwell Scientific Publications.
- CDC (Centres for Disease Control). 1987. International outbreak of type E botulism associated with ungutted, salted white fish. *Californian Morbidity Weekly Reports* 36, 812-3.
- Dodds, K.L. 1993. *Clostridium botulinum* in foods. In "*Clostridium botulinum: Ecology and control in foods*. (Eds. Hauschild, A.H.W. and Dodds, K.L.) pp. 53-68. Inc, New York, Marcel Dekker.
- Dolman, C.E., Dary, G.E. and Lane, R.F. 1961. *Clostridium botulinum* type F with recent observations on other types *J. Infect. Dis.* 109, 107-28.
- Genigeorgis, C. and Riemann, H. 1979. Food Processing and hygiene: In *Food borne infections and intoxications* 2nd edition (Eds. Riemann, H. and Bryan, F.) pp. 613-713, New York Academic Press.
- George Joseph, K., Muraleedharan, V., Kalaimani, N. and Unnikrishnan Nair, T.S. 1986. Quality of cured fish from Tamilnadu coast. *Fish. Technol.* 23(1), 63-5.
- George Joseph, K., Muraleedharan, V., Unnikrishnan, T.S. and Kalaimani, N. 1988. Quality of cured fish from the Maharashtra coast. *Fish Technol.* 25(2), pp. 120-3.
- Gopakumar, K. and Devadasan, K. 1983. Fish curing industry in India. *FAO Report* R.279 Supplement pp. 63-8.
- Haq, I. and Suhadi, F. 1981. Incidence of *Clostridium botulinum* in coastal and inland areas of West Java *Japan J. Med. Sci. Biol.* 34, 231-5.
- Hauschild, A.H.W. 1989 *Clostridium botulinum*. In "*Food-borne bacterial pathogens*" (Ed. Doyle, M.P.) pp. 111-89, New York, Marcel Dekker.

- \_\_\_\_\_. 1993. Epidemiology of human food borne botulism. In "*Clostridium botulinum*: Ecology and control in foods". (Eds. Hauschild, A.H.W. and Dodds, K.L.) pp. 69-104. New York, Marcel Dekker.
- Hayashi, R., Tokuchi, M., Teramoto, K., Fujita, N., Okuno, Y., Rahim, A., Sujudi, R. and Hotta, S. 1981. Distribution of *Clostridium botulinum* in Indonesian soil. In *ICMR Annals* (Ed. Hotta, S.) Vol I. pp. 187-193. Kobe, Int. Cent. For Med. Res. Kobe Univ. Sch. Med.
- Huss, H.H. 1980. Distribution of *Clostridium botulinum*. *Appl. Environ. Microbiol.* 39(4), 764-9.
- \_\_\_\_\_. 1981. *Clostridium botulinum* type E and botulism. 58 pp. Lyngby, Denmark, Technical Univ. of Denmark.
- ICMSF. 1980. *Microbial Ecology of foods* Vol. I. Factors affecting life and death of Microorganisms. International Commission on Microbiological specification of foods, Inc. London, Academic Press.
- IS 2882. 1964. Specification for dried white baits (*Anchoviella* sp.) *Indian Standards Institution*, New Delhi.
- IS 5198. 1969. Specification for dry salted seer fish. *Indian Standards Institution*, New Delhi.
- IS 2883. 1976. Specification for dried white baits. *Indian Standards Institution*, New Delhi.
- IS 594. 1962. Specification for common salt for fish curing (revised). *Indian Standards Institution*, New Delhi.
- Kalaimani, N., Gopakumar, K. and Unnikrishnan Nair, T.S. 1988. Quality characteristics of cured fish of Commerce *Fish Technol.* 25(1), 54-7.
- Lalitha, K.V. and Iyer, K.M. 1990. Isolation of *Clostridium botulinum* from tropical fish. *Letters in Appl. Bacteriol.* 11, 179-81.
- Lalitha, K.V. and P Surendran, P.K. 1993. Isolation of toxigenic strains of *Clostridium botulinum* type D from aquatic environment. In "*Nutrients and bioactive substances in aquatic organisms*". (Eds. Devadasan, K., Mukundan, M.K., Antony, P.D., Viswanathan Nair, P.G., Perigreen P.A. and Jose Joseph). pp. 144-9. Cochin, India, Society of Fisheries Technologists (India).
- \_\_\_\_\_. 1992. Prevalence of *Clostridium botulinum* in fresh retail fish. In "*Proceedings of the fourth Kerala Science Congress*" (Ed. Ramachandran Nair, C.G.) pp. 117-8. Thiruvananthapuram, State Committee on Science, Technology and Environment, Government of Kerala, S.B. Press.
- McClure, P.J., Cole, M.B. and Smelt, J.P.P.M. 1994. Effects of Water activity and pH on growth of *Clostridium botulinum*. *J. Appl. Bacteriol. Symposium Supplement* 76, 105 S-114 S.
- Oguma, K., Yokotta, K., Hayashi, S., Takeshi, K., Kumagai, M., Itoh, N., Tachi, N. and Chiba, S. 1990. Infant botulism due to *Clostridium botulinum* type C toxin. *Lancet* 336, 1449-50.
- Prasad, M.M., Panduranga Rao, C.C. and Gupta, S.S. 1994. Chemical and Microbiological Quality of Dry fish from Kakinada. *Fish. Technol.* 31(1), 75-8.
- Roberts, T.A. and Gibson, A.M. 1979. The relevance of *Clostridium botulinum* type C in public health and food poisoning. *J. Food Technol.* 14(3), 211-66.
- Sakaguchi, G. 1979. Botulism Chapter VIII. In *Food-borne Infections and Intoxications* Second edition (Eds. Riemann, H. and Bryan, F.L.) pp. 389-442. London, Academic Press.

- Slater, P.F., Addiss, D.G., Cohen, A., Levanthal, A., Chassis, G., Zehavi, H., Bashari, A. and Costin, C. 1989. Food borne botulism: an international outbreak. *International J. Epidemiol.* 18, 693-6.
- Smith, G.R. 1990. Botulism. In *Topley and Wilsons Principles of Bacteriology Virology and Immunity*. Vol. III, Bacterial Diseases (Eds. Smith G.R. and Easman, C.S.F. ) pp. 514, 529 London Edward Arnold.
- Solomon, H.M., Rhodehamel, E.J. and Kautter, D.A. 1995. *Clostridium botulinum* Chap. 17. In. *FDA Bacteriological Analytical Manual*. 8<sup>th</sup> edition. pp. 17.01-17.10. AOAC international Gaithersburg MD 20877, USA.
- Sonnabend, O and Sonnabend, W. 1981. Different types of *Clostridium botulinum* (A,D and G) found at autopsy in humans II. Pathological and epidemiological findings in twelve sudden and unexpected deaths. In *Biomedical aspects of botulism* (Eds. Lewis, G.E.Jr.) pp. 303-16. New York Academic Press.
- Sripathy, N.V. 1983. The production and storage from dried fish. *FAO Final Rep. (Supplement)* 279 pp. 1-17.
- Tanasugarn, L. 1979. *Clostridium botulinum* in the Gulf of Thailand *Appl. Environ. Microbiol.* 37(2), 194-7.
- Telzak, E.E., Bell, E.P. and Shultz, S. (1990). An international outbreak of type E botulism due to unviscerated fish. *J. Infect. Dis.* 161, 340-2.
- Troller, J.A. and Christian, J.H.B. 1978. *Water Activity and Food*. p. 131-73. London, Academic Press.
- Weber, J.T., Hibbs, R.G. Jr., Darwish, A., Mishu, B., Corwin, A.L., Rakha, M., Hatheway, C.L., Sharkawy, S.E., El Rahim, S.A., Al-Hamd, M.F.S., Sarn, J.E., Blake, P.A. and Tauxe, R.V. 1993. A massive outbreak of type E botulism associated with traditional salted fish in Cairo. *J. Infect. Dis.*, 167, 451-4.

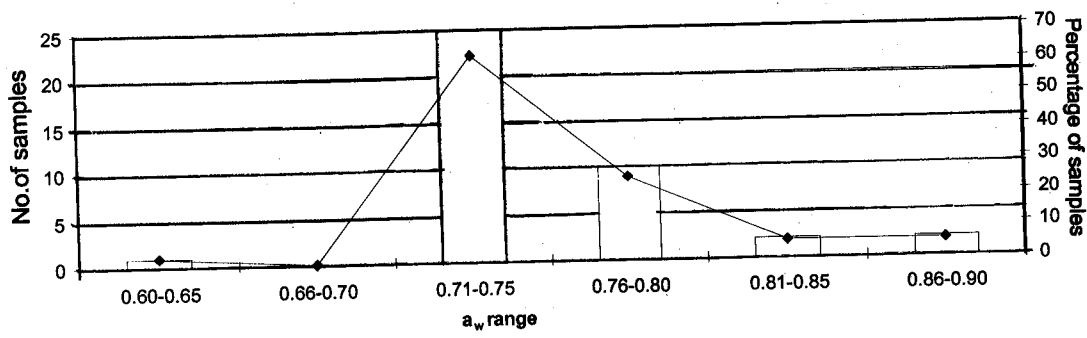


Fig.1.  $a_w$  in cured fish.

□ No. of Samples    ◆ % of Samples

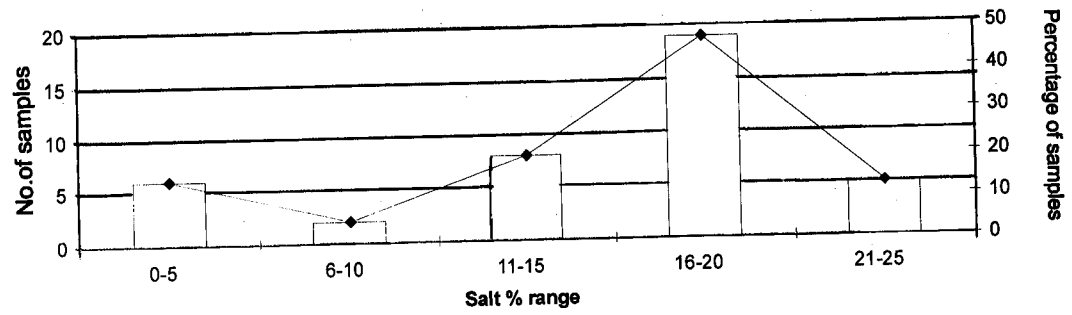


Fig.2. Percentage of salt in cured fish.

□ No. of Samples    ◆ % of Samples in each salt range

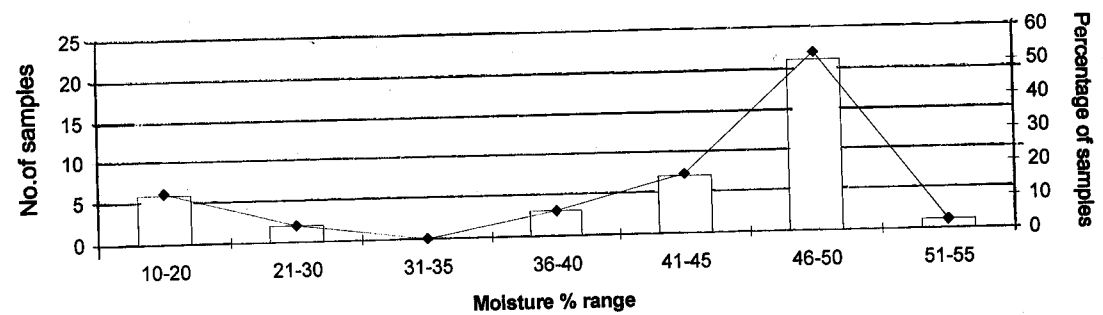


Fig.3. Percentage of moisture in cured fish.

□ No. of Samples    ◆ % of Samples